



Oral microbiome, COVID-19 and probiotics

Oralni mikrobiom, COVID-19 i probiotici

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Introduction

The COVID-19 pandemic is an ongoing global pandemic that seriously endangers human life and health. Clinical presentation of this disease varies from completely asymptomatic or mild infection to severe complications, post-COVID syndrome, and even lethal outcome¹.

The etiologic agent of COVID-19 disease is the SARS-CoV-2 virus, an RNA-positive single-stranded virus from the *Coronaviridae* family. Even though coronaviruses primarily cause zoonotic infections, there are currently seven strains that can cause an infection in humans². There are five different variants resulting from genetic evolution that have been identified since the onset of the pandemic: Alpha (B.1.1.7), Beta (B.1.351), Gamma (P.1), Delta (B.1.617.2), and Omicron (B.1.1.529)³.

The SARS-CoV-2 virus is one of many respiratory viruses where the oropharynx is the primary site of entry and replication⁴. It binds to angiotensin-converting enzyme 2 (ACE-2) receptors *via* its glycoprotein extension (S protein)⁴⁻⁶. These receptors are present on the tongue, oral and nasal mucosa, salivary glands, and nasopharynx⁷⁻¹¹. The study of Xu et al.¹² showed higher ACE-2 expression in small salivary glands than in lungs during COVID-19 disease, proving that salivary glands represent a significant virus reservoir¹³.

Since the beginning of the COVID-19 pandemic, basic preventive measures have been applied to prevent the transmission of the virus (hand disinfection, wearing face masks, social distancing, and quarantine). Thirteen different official treatment protocols were introduced, though currently, there is no optimal treatment¹. Antiviral therapy, vaccines, and immunomodulating agents are widely used in

order to reduce disease severity, especially in patients with an increased risk of developing a severe clinical form of the disease¹⁴. While searching for the most effective treatment against the COVID-19 infection, scientists and clinicians have also applied alternative possibilities for improving immunity¹⁵. Recent research data have revealed the interaction of intestinal and respiratory systems in immunity induction, acting as local and systematic modulators of inflammation¹⁶.

The human microbiome is important for blocking inflammation and immunity regulation. The impact of oral microbiome (OM) dysbiosis in patients with COVID-19, which can directly or indirectly favor the development of the infection and affect the pathogenesis of the disease, has been recognized. The human microbiome plays a significant role in the immune response of the host to respiratory viral infections. The modulation of local and systemic immune reactions by using probiotics is one of the most promising effects of probiotics on overall human health¹⁵. The presence and registration of dominant microbial communities in an individual's OM can enable personalized therapy that aims at restoring the microbiome and preventing the occurrence of many diseases in the future¹⁷.

Many studies on COVID-19 published in the past three years have examined the exact mechanism of virus replication at the primary site of infection, as well as the role of OM on SARS-CoV-2 virus binding capacity and infection development^{4, 5, 9, 18-25}. The goal of this review was to evaluate the role of OM in the prevention of SARS-Cov-2 virus infection and its impact on the severity of COVID-19 clinical presentation. In addition, the aim of this literature review was to present possible preventive and therapeutic applications of probiotics which were used as one of the

remedies against SARS-Cov-2 virus infection. This review focuses on the analysis of oral microbiota during the COVID-19 infection and gives us new insights into the relationship between microbiota and probiotics.

Oral microbiome

The human OM is the genome of all microorganisms which inhabit the oral cavity. The term “microbiota” refers to a specific and unique composition of microbial population that affects health and varies from person to person. In many studies, these two terms are equated^{26–28}.

Oral flora is the second largest and one of the most diverse microbiomes in the human body, right after the intestines, which weigh about 2 kg^{29–32}. The composition of oral flora is heterogeneous and contains over 1,000 different bacterial species, viruses, fungi, helminths, protozoa, and archaea that persist in mutual balance but also in symbiosis with the host^{23, 33–35}. Each person has a complex of microorganisms, and everyone carries an individual microbiome that develops over a lifetime. The composition and diversity of the OM can be influenced by the following: the duration of pregnancy, delivery method, breastfeeding, genetic factors (sibling microbiota profiles are more similar when compared to the profiles of persons who are not related), environmental conditions (oral hygiene, saliva quality, and quantity), habits (tobacco, alcohol, stress, etc.), diet, certain drugs (antibiotics, antacids, etc.), age (three stages of evolutionary development: childhood, adulthood, and old age), and individual general health^{4, 36–42}.

The most common species of bacteria in the OM are representatives of the genera *Bacteroides*, *Synergistes*, *Gemella*, *Granulicatella*, *Streptococcus*, *Veillonella*, the phyla Actinobacteria, Proteobacteria, Tenericutes, Firmicutes, and

Spirochaetes, while oral viruses are mainly composed of eukaryotic viruses such as *Herpesviridae*, *Papillomaviridae*, and *Anelloviridae* [human papillomavirus (HPV), human cytomegalovirus (CMV), herpes simplex virus type-1 (HSV-1), and Epstein-Barr virus (EBV)]^{32, 43}. *Myoviridae* and *Podoviridae* belong to lytic viruses (they rapidly degrade their bacterial hosts), while *Siphoviridae* are lysogenic viruses that are in balance with the host bacteria. Oral viruses represent a restricting factor for bacterial growth and can control bacterial oral populations^{44, 45}. Research by Peters et al.⁴⁶ described 154 species of commensal fungi and confirmed that the *Candida* genus was most commonly present in 70% of healthy patients. Recent studies revealed the commensal presence of different genera of protozoa, helminths, and archaea. Nonetheless, their pathogenic potential in the development of oral diseases is still unexplored^{47–49}.

Importance of oral microbiome in oral and general health

The oropharyngeal microflora in a healthy host maintains balanced symbiotic relationships defined as “microbial homeostasis” (eubiosis)³². The OM is exposed to frequent daily fluctuations that can lead to microbial imbalance (dysbiosis)^{50, 51}. Microbial balance in the oral cavity is necessary because it enables equilibrium between beneficial and harmful microorganisms that interact with each other and can have an inhibitory, stimulating, or synergistic effect on each other^{32, 50, 51}. The OM contributes to the development of the local immune system. However, its imbalance, along with the complex interaction with the host resistance and various environmental factors, creates conditions for the development of various oral or systemic diseases (Figure 1)^{17, 52, 53}. During dysbiosis, the following three changes occur: loss of

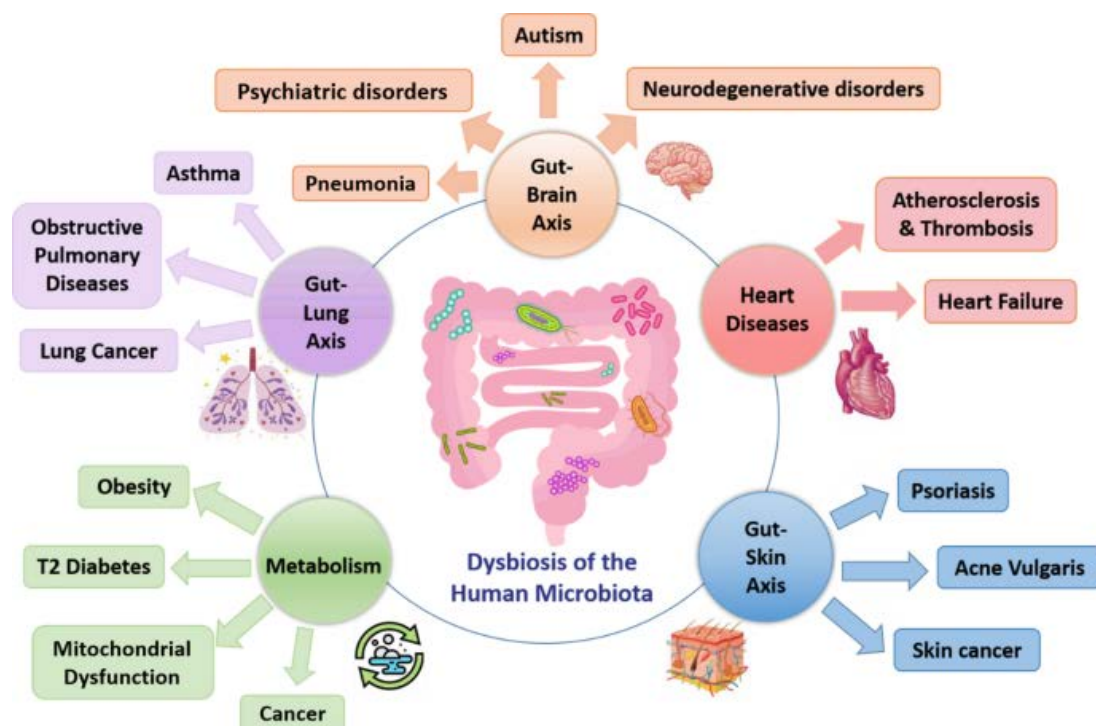


Fig. 1 - Impact of microbiota dysbiosis on the overall health of humans⁵².

microbial diversity, loss of beneficial microorganisms, and increase in pathogenic microorganisms that are not mutually exclusive and can occur simultaneously⁵⁴. As a result, potentiating cariogenic and inflammatory bacterial genomes^{33, 55} make perfect conditions for the development of diseases such as caries⁵⁶⁻⁵⁹, periodontal diseases⁶⁰⁻⁶⁴, and malignant alterations of oral tissues⁶⁵⁻⁶⁷.

Many oral microorganisms enter the digestive tract through saliva, thus changing the intestinal microbiome, which plays an important role in digestion and absorption of nutrients, formation of a protective barrier against pathogens, development and regulation of the immune system, enzyme and vitamin production, control of inflammatory reactions, and neuro and psychological regulation³⁵. Consequently, the connection between the oropharyngeal and intestinal microbiome sheds new light on inflammatory processes and their therapy. The link between oral and general health is well documented in the literature^{35, 50, 52, 68-76}. Chronic oral infections lead to the release of proinflammatory mediators, i.e., cytokines that further propagate inflammatory processes and increase the risk of muscle and digestive problems^{61, 72}, bronchopulmonary diseases^{37, 73}, rheumatoid arthritis (RA)^{53, 67}, complications during pregnancy and childbirth^{38, 41, 64, 71, 75}, cardiovascular diseases^{61, 67, 69, 70, 75, 76}, obesity⁴², liver disease⁶⁷, oral cancer⁶⁵, pancreatic cancer⁷⁷, type 2 diabetes^{67, 68}, Parkinson's disease⁷⁴, psychiatric disorders⁷⁸, and colorectal cancer⁶⁷. Each disease is characterized by unique oral and intestinal microbial changes⁷⁹. Microbiome rebalancing is closely related to the recovery from the primary disease, which proves the significant role of the microbiome in healing^{72, 80}.

Maintaining good oral hygiene is, therefore, important in controlling oral bacterial status, maintaining or restoring symbiotic homeostasis, as well as preventing the spread of oral pathogens to other parts of the body⁸¹⁻⁸³.

Disruption of the oral microbiome during and after SARS-CoV-2 virus infection

Human OM can have a great influence on the regulation of innate and acquired immunity to viral infections⁴, which is especially important for viruses that enter the body through the oropharynx^{5, 23, 84-86}. Aerosol respiratory viruses encounter the OM of the upper respiratory tract and favor its dysbiosis and disease progression^{87, 88} (e.g., the microbiome in patients with influenza is characterized by the abundance of genus *Pseudomonas*)⁷⁵. Oral dysbiosis promotes respiratory infections directly by increasing pathogenic bacteria and aspirating oral pathogens into the respiratory organs. Indirectly, it alters the immune response of respiratory epithelium and promotes the adhesion of pathogens, cytokine secretion, and production of enzymes that interfere with pathogen clearance^{23, 89}. Even in the case of SARS-CoV-2 virus infection, oral dysbiosis may favor the development of infection by these mechanisms⁴. On the other hand, OM can also contribute to the regulation of immunity and inflammation blockade^{13, 30, 84, 90}. The study of Pfeiffer and Sonnenburg⁹¹ showed that oral microbiota commensals could also produce

antiviral compounds (defensins) against several viral genera (*Adenovirus*, *Herpesvirus*, *Papillomavirus*, *Orthomyxovirus*, and *Coronavirus*).

The oropharyngeal and intestinal microbiota have the possibility of co-infection with microorganisms originating from the oral cavity. The correlation between the imbalance of OM and the increased number of dysbiotic species may serve as predictive factors for COVID-19 disease^{5, 35, 79, 92-104}. Ward et al.²⁴ showed that the severity of COVID-19 disease could be predicted according to the composition of the intestinal or OM. Two pathogens, *Porphyromonas endodontalis* in oral and *Enterococcus faecalis* in the intestinal microbiome, may serve as a predictor of the severity of SARS-CoV-2 infection. The authors suggested that the key to prioritizing patients who need urgent treatment is the identification of biomarkers that can predict clinical outcomes of COVID-19 disease²⁴.

Haran et al.²³ described how the dysbiosis of the inflammatory type of OM, characterized by the members of genera *Prevotella* and *Veillonella*, may play an important role in prolonging the duration of COVID-19 symptoms. The genus *Veillonella* are gram-negative anaerobic cocci that produce large amounts of lipopolysaccharides and may be an additional co-infectious agent (especially *Veillonella dispar* and *Veillonella infantium*)⁹⁸. The genus *Prevotella* is highly inflammatory and affects the promotion of SARS-CoV-2 infection, thus worsening the severity of the COVID-19 clinical presentation¹⁰⁵⁻¹⁰⁷. Haran et al.²³ emphasized the importance of the presence of gram-negative bacteria with a liposaccharide component in the capsule (*Leptotrichia* and various species of genus *Veillonella*) which can have proinflammatory effects and cause systemic damage and neuroinflammation. Due to OM dysbiosis and the presence of pathogens that promote chronic inflammation (*Leptotrichia*, *Prevotella*, and *Fusobacterium*), myalgic encephalomyelitis and symptoms of neurological diseases (confusion, disorientation, slow thinking, and poor concentration after six months) may occur in COVID-19¹⁰²⁻¹⁰⁴. The present commensals (*Prevotella* and *Neisseria*) in OM can act as a local probiotic and counteract the SARS-CoV-2 virus¹⁷. Likewise, the higher relative abundance of the *Rothia* genus in the patient's oral flora affects the occurrence of COVID-19 complications¹⁰⁸.

Ren et al.⁷⁹ identified specific microbial markers of oral microbiota in patients with COVID-19 but also in recovered patients. The results of this study showed compositional and functional changes in OM of COVID-19 patients. During infection, there is an overall decrease in the diversity of oral microorganisms. The number of bacteria that produce lipopolysaccharides was increased, while the number of bacteria that produce butyric acid was decreased. Thus, by secreting lipids into the bloodstream, microbiome dysbiosis may affect the progression of COVID-19. Ren et al.⁷⁹ also noticed a better prognosis in patients with severe COVID-19 who had an OM enriched with *Streptococcus* (*S. parasanguinis*). Oral dysbiosis persisted even after the recovery from COVID-19 infection when a constant increase in *Porphyromonas* and *Haemophilus* gen-

era and a decrease in *Leptotrichia*, *Megasphaera*, and *Selenomonas* (*Megasphaera* is a cariogenic bacterium) ¹⁰⁶ genera was detected.

Furthermore, an imbalance in the relative numbers of different bacterial strains and the genera *Enterococcus* and *Enterobacter* were present only in patients with COVID-19 (not observed in the control group of healthy patients) ⁵. Ward et al. ²⁴ showed that higher quantities of *Porphyromonas endodontalis* are correlated with an increase in severe stages of COVID-19, while higher quantities of *Muribaculum intestinale* are linked with moderate cases. Cox et al. ⁹⁵ also highlighted the impact of coinfections on the clinical presentation and mortality of patients with COVID-19. They indicated an association between cariogenic and oral pathogenic bacteria and complications of COVID-19. According to the earlier literature evidence, these bacteria are involved in the pathogenesis of respiratory and chronic inflammatory systemic diseases (type 2 diabetes mellitus, hypertension, cardiovascular disease), which are also the most common comorbidities associated with the risk of severe complications and death from COVID-19 ^{97, 109, 110}. Marouf et al. ¹⁰¹ also observed an abundance of periodontopathogenic bacteria in COVID-19 patients and demonstrated that the presence of periodontitis was associated with a 3.5-fold higher risk of admission to intensive care units, a 4.5-fold higher risk of assisted ventilation, and an 8.81-fold higher risk of mortality independent of other concomitant risk factors. On the other hand, numerous studies reported that interventions aimed at boosting oral hygiene in patients with pneumonia have significantly improved the clinical picture and reduced mortality ^{23, 35, 105, 107}.

The exact genome of oral flora in patients with COVID-19 is still the focus of scientific interest. Iebba et al. ²⁰ were among the first who had described the bacterial component of OM in patients with COVID-19, pointing to the importance of fungi and viruses in defining individual sensitivity. By examining an oropharyngeal swab, Ai et al. ⁹³ found that more than half of COVID-19 patients had co-infection with another virus, such as influenza A or B, rhinoviruses, enteroviruses, or respiratory syncytial virus. Soffritti et al. ⁵ investigated an association between OM profile and severity of COVID-19 clinical presentation and observed a significant increase in *Herpesviridae* viruses, EBV, and HSV-1. EBV infection in patients with COVID-19 has been associated with an increased risk of severe COVID-19 symptoms as well as a fatal outcome ^{96, 99}.

Changes have also been observed in the fungal part of the microbiome, with the appearance of *Aspergillus*, *Nakaseomices*, and *Malassezia spp.*, *Candida albicans*, *Saccharomyces cerevisiae*, *Aspergillus fumigatus*, and *Malassezia restricta* (OM of healthy patients consists only of *Candida* and *Candida cerevisiae*, *Aspergillus fumigatus*, and *Malassezia restricta*) ⁵. According to Jasinski-Bergner et al. ¹⁰⁹, these changes in COVID-19 patients could be an inducing factor influencing the onset of SARS-CoV-2 virus infection because dysbiosis facilitates the activation or reactivation of oral pathogens, which can further impair proper immune

control and lead to deterioration of immune response effectiveness.

Ward et al. ²⁴ found that the composition of OM has a high accuracy of COVID-19 severity prediction (84% accuracy of predicting fatal outcome).

Application of probiotics

With the emergence of increasing antibiotic resistance, new concepts for the prevention and therapy of multidrug-resistant infections are proposed by causing the microbiological shift of the endogenous microbiota. For this reason, research into bactericidal action and antiviral factors of probiotics became the focus of modern interest ^{107, 111–113}.

Positive effects of probiotics are primarily observed in the treatment of gastrointestinal infections but also in the prevention of various pathological conditions in the field of gastroenterology, allergology, internal medicine, oncology, oral medicine, pediatrics, infectiology, and psychiatry ^{104–107, 109–114}.

Modern therapeutic procedures have introduced changes in treatment protocols with a tendency to establish a healthy environment to prevent the development of opportunistic infections and recover the OM. Probiotics are living microorganisms (so-called “good” bacteria) that, if applied in an adequate amount, could establish and maintain the ecological balance of microflora. They are safe, non-pathogenic, non-invasive, and non-carcinogenic strains that can perform recolonization and restore symbiosis between the host and the disturbed microbiota ¹¹⁵. Several bacterial genera are most often used as probiotics: *Bifidobacterium*, *Lactobacillus*, *Bacillus*, and *Pediococcus*. The strains used most often are *Bifidobacterium bifidum*, *Bifidobacterium breve*, *Bifidobacterium infantis*, *Bifidobacterium longum*, *Lactobacillus acidophilus*, *Lactobacillus casei*, *Lactobacillus plantarum*, *Lactobacillus reuteri*, *Lactobacillus rhamnosus*. In addition, fungi can be used as probiotics ^{112–115}.

Probiotics act in complex multifactorial ways. Probiotic bacteria can interfere with the absorption process by directly binding to the virus or inhibiting the entry into the epithelial cells by blocking the host receptor ²⁵. Probiotics can also compete with nutrient pathogens, produce antimicrobial agents, strengthen the intestinal epithelial barrier, and modulate the immune system of the host ^{116–118}. The performance of probiotics can be twofold – immunostimulatory and immunoregulatory. A group of immunostimulatory probiotics affects the proliferation of T helper (Th) 1 cells and stimulates the production of interleukin (IL)-12, which induces the production of interferon-gamma (IFN- γ) in natural killer cells. Immunoregulatory probiotics stimulate regulatory T cells but also suppress proinflammatory responses by inducing IL-10 ¹¹⁷. Experimental animal models showed that balancing cellular and humoral immune responses mitigates the effects of a “cytokine storm” ^{25, 119}.

The significant role of probiotics in maintaining homeostasis of the upper respiratory tract microbiome was proven in multiple studies. In addition, it was revealed that oropharyngeal probiotics are very effective in maintaining immune

system stability and protecting against viral infections^{120–122}. Direct and indirect efficiency of various probiotic strains (e.g., *Lactococcus lactis* JCM 5805 and *Bacteroides breve* IIT4064) has been proven against influenza virus¹²³. Probiotic bacteria release various substances, such as bacteriocins, biosurfactants, lactic acid, hydrogen peroxide, nitric oxide, and organic acids, which can inhibit virus proliferation²⁵. *Lactobacillus* genus produces lactic acid as an antiviral inhibitory metabolite, thus preventing secondary infections¹²⁴. Furthermore, like the genus *Bifidobacterium*, *Lactobacillus* genus can capture the virus and interfere with the binding of the virus to the receptors of the host cell^{125, 126}. Nisin and peptide P18 are bacteriocins with antiviral effects against influenza A virus¹²⁷. Apart from bacteriocin production, the antiviral ability of oropharyngeal probiotics is also maintained by the stimulatory effect on the innate immune response, which is manifested by an increase in the IFN- γ levels in human saliva ten hours after oral administration of *Streptococcus salivarius* (strain K12) lozenge¹²⁸. Probiotic strains of genera *Lactobacillus* and *Bifidobacterium* (such as *Lactobacillus reuteri* ATCC 55730, *Lactobacillus paracasei*, *Lactobacillus casei* 431, *Lactobacillus fermentum* PCC, and *Bifidobacterium infantis* 35624) are significant factors in generating immunomodulatory responses during various infections^{25, 129}. In addition, probiotic bacteria have antioxidant potential in neutralizing free radicals. The strains *Lactobacillus rhamnosus* GG, *Lactobacillus plantarum* CAI6, *Clostridium butyricum* MIIAIRI 588, and strains in VSL#3[®] increase total antioxidant capacity¹³⁰. Moreover, by participating in the formation of redox homeostasis, probiotics can inhibit the progression of COVID-19 disease²⁵.

Probiotics and COVID-19

The proven effectiveness of probiotics both in the treatment and prevention of viral infections was the rationale behind their use in patients with SARS-CoV-2 virus infection^{25, 28, 131–135}.

In the last three years, studies dealing with the importance and benefits of probiotics in the prevention and treatment of COVID-19 have been conducted (Table 1)^{21, 135–143}. The use of probiotics, along with other therapies, led to a shorter and easier clinical presentation, with reduced severity of gastrointestinal and respiratory symptoms, a lower percentage of smell and taste disorders, and less frequent symptoms of post-COVID syndrome^{125–143}. However, in patients on corticosteroid therapy, probiotic supplemental therapy is contraindicated due to their primary disease¹³⁵.

The first study that proved the positive effects of probiotics in the treatment of COVID-19 infection was the 2020 Wuhan study²¹. The use of the probiotic strain ENT-K12 (*Streptococcus thermophilus*) among medical workers in institutions for COVID-19 treatment has reduced the possibility of respiratory infection and lethal outcomes by 80%. This probiotic strain locally releases two antibiotics (salivaricin A2 and B) and reduces the possibility of colonization of β -hemolytic group A streptococci, including *Streptococcus pyogenes* (a bacterial pathogen that causes coinfection during

viral infection). The use of probiotics has also reduced the use of antibiotics among the respondents by more than 90%²¹.

Block¹⁴⁴ suggested the concomitant use of probiotics in patients with COVID-19, treated with azithromycin, to reduce the risk of hypercolonization of *Candida albicans* strains. Nutritional support with probiotic strains of *Lactobacillus acidophilus*, *Bifidobacterium*, and *Saccharomyces boulardii*, along with minerals and vitamins, has reduced the complications of massive antibiotic therapy^{145–147}.

D'Ettorre et al.¹³⁵ compared the incidence of respiratory failure and control of symptoms, after probiotic therapy, with different *Streptococcus*, *Lactobacillus*, and *Bifidobacterium* strains. The use of probiotics was associated with a lower risk of respiratory failure and faster control of COVID-19 symptoms (especially diarrhea). In patients with a severe clinical picture, the immunomodulatory effects of probiotics may be relevant for the prevention of acute respiratory distress syndrome and multiple organ failure as a complication of cytokine storm¹³⁵.

Ceccarelli et al.¹⁴⁷ observed a lower mortality rate after the use of probiotic strains of genera *Streptococcus*, *Bifidobacterium*, and *Lactobacillus* during COVID-19, but with longer hospital stays. However, research by Bozkurt and Bilen¹⁴² showed that in patients with moderate and severe COVID-19 symptoms, an additional therapeutic dose of the probiotic strain *Bifidobacterium animalis* resulted in a shorter hospital stay and lower mortality rates.

Probiotics mechanism of action in COVID-19

The mechanism of probiotic protection against SARS-CoV-2 infections is based on general effective principles, such as inhibition of pathogen adhesion and antimicrobial and immunomodulatory specific properties of different probiotic strains^{118, 145, 148}. These mechanisms can enhance the elimination of the SARS-CoV-2 virus but also act preventively by suppressing bacterial coinfections that correlate with COVID-19 (Figure 2)^{118, 148, 149}.

During fermentation, probiotic strains produce specific bioactive peptides that block ACE-2 enzyme receptors, thus preventing the SARS-CoV-2 virus from binding to these active sites^{4, 150, 151}. The remnants of dead probiotic cells can act as ACE-2 inhibitors as well. These bioactive peptides may modulate blood pressure due to the inhibition of the conversion of angiotensin-I to angiotensin-II. The possible effect of these peptides on reducing the progression of COVID-19 is still being examined¹⁵². The concept of using ACE-2 receptor-blocking drugs as a treatment modality for COVID-19 was first proposed by Fernandez-Fernandez¹⁵¹. Imai et al.¹⁵² reported that the usage of ACE blockers had a positive effect on the reduction of respiratory distress syndrome. The study by Singh and Rao²⁵ confirmed that by binding mucosal cell receptor and ACE-2, probiotic strains interfere with coronavirus and block its binding, while an increase of innate immunity is stimulated by releasing intestinal mucins from mucosal cells and producing secretory antibodies (IgA). The authors stated that the key role in combati-

Table 1 Review of clinical studies which used probiotics in the prevention and therapy of COVID-19 disease *

Reference	Country	Study type	Subjects	Probiotic strain	Intervention	Main results
d'Ettore et al. ¹³⁵	Italy	Single group	70 patients with COVID-19 hospitalized	<i>Streptococcus thermophilus</i> DSM 32345, <i>Lactobacillus acidophilus</i> DSM 32241, <i>Lactobacillus helveticus</i> DSM 32242, <i>Lactocaseibacillus paracasei</i> DSM 32243, <i>Lactiplantibacillus plantarum</i> DSM 32244, <i>Levilactobacillus brevis</i> DSM 27961, <i>Bifidobacterium lactis</i> DSM 32246, <i>Bifidobacterium lactis</i> DSM 32247	Daily oral 2.4 billion CFUs bacteria for a period of 14 days	Probiotic intervention demonstrated a significant improvement in clinical conditions among patients with COVID-19.
Tang et al. ¹³⁶	The United States	Double-blinded, randomized, placebo-controlled trial	1,132 individuals with household contacts who tested positive for COVID-19	<i>Lactocaseibacillus rhamnosus</i> GG (ATCC 53103)	Daily oral <i>Lactocaseibacillus rhamnosus</i> GG or placebo for a period of 28 days	Low-cost and safe probiotics can serve as a rapid intervention strategy in the prevention or reduction of symptoms of pandemic diseases.
Endam et al. ¹³⁷	Canada, Saudi Arabia, and the United States	Prospective randomized clinical trial	23 individuals between 18–59 years of age received late PCR positive tests for SARS-CoV-2	<i>Lactococcus lactis</i> W136	Nasal irrigations through a buffered isotonic solution containing 2.4×10^9 CFUs of <i>Lactococcus lactis</i> W136 or buffered isotonic saline isolated for two weeks (twice a day)	Probiotic intranasal intervention was correlated with a reduced number of patients showing moderate/severe symptoms of fatigue, loss of smell perception, and sensation of breathlessness, and by decreased percentage of individuals with moderate/severe facial pain or sore throat.
Gutierrez-Castrellon et al. ¹³⁸	Mexico and Spain	Single-center, quadruple-blinded randomized clinical trial	300 outpatients with symptomatic COVID-19 (ages 18–60) with positive nucleic acids test for SARS-CoV-2	<i>Lactiplantibacillus plantarum</i> KABP022, KABP023 and KABP033, <i>Pediococcus acidilactici</i> KABP021	Daily ingestion of 10^9 CFUs for a period of 30 days	Remission was achieved by 53% of probiotic group compared to 28% in placebo group.
Mozota et al. ¹³⁹	Spain	Single group	29 residents of a nursing home who tested positive for COVID-19	<i>Ligilactobacillus salivarius</i> MP101	Daily consumption of 10^9 CFUs of <i>Ligilactobacillus salivarius</i> MP101 per unit of product (125 g)	Certain immune factors can be utilized as possible nasal or fecal biomarkers of benefits of probiotic strain supplementation in the diet of elderly people infected with SARS-CoV-2.
Wang et al. ²¹	China	Retrospective study	138 patients	5×10^7 CFUs of live <i>Bifidobacterium longum</i> ; live <i>Lactobacillus bulgaricus</i> and <i>Streptococcus thermophilus</i> (should not be lower than 0.5×10^6 CFUs)	Four doses at a time, 3 times a day	Compared to the control group, patients treated with probiotics showed a significantly reduced time for achieving a negative nucleic acid test while the inflammation indexes, including PCT and CRP, were significantly reduced.
Wang et al. ¹⁴⁰	China	Randomized controlled clinical trial	200 frontline medical staff	One billion CFUs of <i>Streptococcus thermophilus</i> ENT-K12 over shelf-life	Slowly dissolving oral lozenge	Significantly reduced incidence of respiratory tract infections by 64.8%, reduced the time experiencing respiratory tract infections and oral ulcer symptoms

Table 1 (continued)

Wang et al. ¹⁴⁰	China	Randomized controlled clinical trial	200 frontline medical staff	One billion CFUs of <i>Streptococcus thermophilus</i> ENT-K12 over shelf-life	Slowly dissolving oral lozenge	Significantly reduced incidence of respiratory tract infections by 64.8%, reduced the time experiencing respiratory tract infections and oral ulcer symptoms by 78%, shortened the sick-leave days by 95.5%, and reduced the time under medication in cases when there was no record of antibiotic and anti-viral drug intake in the probiotic group.
Li et al. ¹⁴¹	Wuhan, China	Retrospective single-center study	311 COVID-19 patients	<i>Lactobacillus rhamnosus</i> GG	Daily oral administration per 32 days	Moderates immunity and decreases the incidence of secondary infection in COVID-19 patients.
Bozkurt and Bilen ¹⁴²	Turkey	Retrospective study	44 moderate/severely ill adults	One trillion CFUs <i>Bifidobacterium</i> BB-12 strain	Oral administration in 250 mL water. Total doses were divided into three parts and administered to patients for 3 days	Lower mortality, shortening the length of stay in hospital, early radiologic improvement and decrease plasma IL-6 level in moderate/severe SARS-CoV-2 patients in the probiotic group.
Wischmeyer et al. ¹⁴³	The United States	Randomized, double-blind, placebo-controlled trial	182 participants	<i>Lactobacillus rhamnosus</i> GG	Daily oral use for 28 days	Prolonged time for development of COVID-19 infection, reduced incidence of symptoms and changes to gut microbiome structure when used as post-exposure prophylaxis within seven days after probiotic exposure.

COVID-19 – corona virus disease-19; CFUs – colony forming units; PCR – polymerase chain reaction test; CRP – c-reactive protein; PCT – procalcitonin; IL – interleukin; SARS-CoV-2 – severe acute respiratory syndrome coronavirus 2.

*Modified Table of Xavier-Santos et al. ¹³⁴

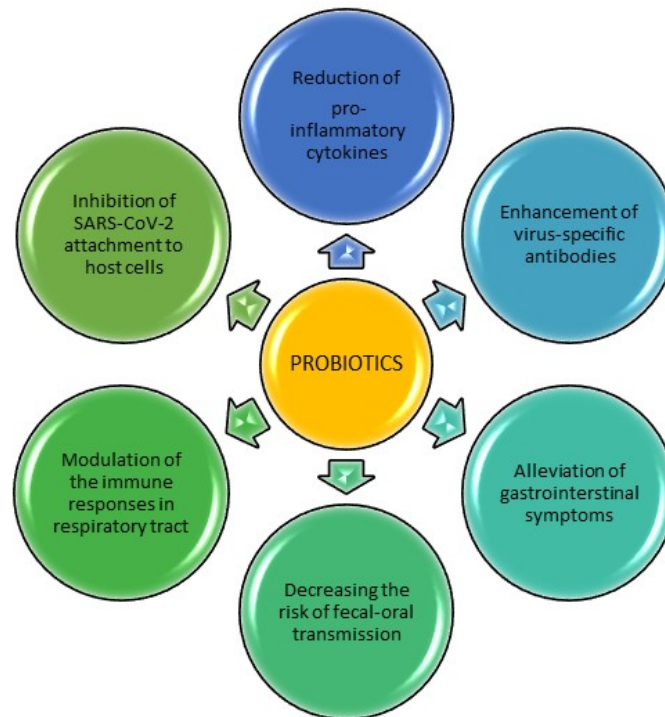


Fig. 2 - Antiviral mechanisms of probiotics against SARS-CoV-2 infection.

ng coronavirus proliferation is played by modulation of immune responses and balance of acquired immunity, production of inflammatory cytokines, the proliferation of B cells that produce specific antibodies, and activation of cytotoxic T lymphocytes that participate in the adaptive immune response²⁷. Baidara et al.¹²⁶ also showed that some probiotics improved the regulatory activity of T cells and reduced the production of proinflammatory cytokines. In addition, the antiviral effect of probiotics may be achieved by a large number of secreted specific metabolites and bacteriocins¹⁵³.

Strengthening the immune response during incubation and the initial phase of COVID-19 disease is crucial in eliminating the virus and preventing the progression of the disease. The use of certain strains of *Bifidobacterium* or *Lactobacillus* has a great influence on the elimination of the SARS-CoV-2 virus from respiratory organs¹⁵⁴. The use of probiotics, along with adequate treatment for COVID-19, can significantly reduce the occurrence and duration of various systemic diseases^{154, 155}.

The limitations of this comprehensive literature review arise due to the high heterogeneity of studies that investigated the change and impact of oral microbiota during COVID-19 without knowing the previous status of patients' OM and because patients with different immune statuses used different probiotic strains.

It should be emphasized that introducing targeted drugs and beneficial bacteria was of great importance in restoring

the damaged microbiome. Further research should be directed to discovering the most effective probiotic strains, doses, and formulations, as well as the interaction of probiotics and microbiomes. In addition, the influence of environmental factors on the oropharyngeal microbiome, as well as possible coinfection, should be further investigated.

Conclusion

After an extensive review of the literature, it was concluded that numerous clinical studies showed that OM might influence resistance to primary infection and be a predictor for disease severity and complications during COVID-19. The use of probiotic strains can inhibit the adhesion of pathogens, improve the barrier function of the intestine and strengthen the immune response. Through these mechanisms, probiotics can reduce the progression and the development of more severe forms of the disease, shorten the hospital stay and reduce the frequency of post-COVID syndrome.

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R E F E R E N C E S

- World health organization (WHO). WHO coronavirus (COVID-19) dashboard. [Internet] 2022. Available from: <https://covid19.who.int/> [Accessed on 22 Apr 2022].
- Ye ZW, Yuan S, Yuen KS, Fung SY, Chan CP, Jin DY. Zoonotic origins of human coronaviruses. *Int J Biol Sci* 2020; 16(10): 1686–97.
- Łoniewski I, Skonieczna-Żydecka K, Solek-Pastuszka J, Marlicz W. Probiotics in the management of mental and gastrointestinal post-COVID symptoms. *J Clin Med* 2022; 11(17): 5155.
- Baghbani T, Nikzad H, Azadbakht J, Izadpanah F, Haddad Kasbani H. Dual and mutual interaction between microbiota and viral infections: a possible treat for COVID-19. *Microb Cell Fact* 2020; 19(1): 217.
- Soffritti I, D'Acolti M, Fabbri C, Passaro A, Manfredini R, Zuliani G, et al. Oral microbiome dysbiosis is associated with symptoms severity and local immune/inflammatory response in COVID-19 patients: a cross-sectional study. *Front Microbiol* 2021; 12: 687513.
- Bobórz-Ávila S, Bernal-Cepeda L, Reina-Marin M, Navarro-Saiz L, Castellanos J. The mouth, oral health, and infection with SARS-CoV-2: an underestimated topic. *Infectio* 2022; 26 (1): 78–82.
- To KK, Tsang OT, Yip CC, Chan KH, Wu TC, Chan JM, et al. Consistent detection of 2019 novel coronavirus in saliva. *Clin Infect Dis* 2020; 71(15): 841–3.
- Amorim Dos Santos J, Normando AGC, Carvalho da Silva RL, De Paula RM, Cembranel AC, Santos-Silva AR, et al. Oral mucosal lesions in a COVID-19 patient: new signs or secondary manifestations? *Int J Infect Dis* 2020; 97: 326–8.
- Bao L, Zhang C, Dong J, Zhao L, Li Y, Sun J. Oral microbiome and SARS-CoV-2: beware of lung co-infection. *Front Microbiol* 2020; 11: 1840.
- Silva A, Azevedo M, Sampaio-Maia B, Sousa-Pinto B. The effect of mouthrinses on severe acute respiratory syndrome coronavirus 2 viral load: a systematic review. *J Am Dent Assoc* 2022: 635–48. [Epub ahead of print].
- Paradowska-Stolarz AM. Oral manifestations of COVID-19: brief review. *Dent Med Probl* 2021; 58(1): 123–6.
- Xu J, Li Y, Gan F, Du Y, Yao Y. Salivary glands: Potential reservoirs for COVID-19 asymptomatic infection. *J Dent Res* 2020; 99(8): 989.
- Mohapatra RK, Dhama K, Mishra S, Sarangi AK, Kandi V, Tiwari R, et al. The microbiota-related coinfections in COVID-19 patients: a real challenge. *Beni Suef Univ J Basic Appl Sci* 2021; 10 (1): 47.
- Rocchi G, Giovanetti M, Benedetti F, Borsetti A, Ceccarelli G, Zella D, et al. Gut microbiota and COVID-19: potential implications for disease severity. *Pathogens* 2022; 11(9): 1050.
- Amrouche T, Chikinda ML. Probiotics for immunomodulation in prevention against respiratory viral infections with special emphasis on COVID-19. *AIMS Microbiol* 2022; 8(3): 338–56.
- Cyprian F, Sobail MU, Abdelhafez I, Salman S, Attique Z, Kamareddine L, et al. SARS-CoV-2 and immune-microbiome interactions: lessons from respiratory viral infections. *Int J Infect Dis* 2021; 105: 540–50.
- Rafiqul Islam SM, Foyzal MJ, Hoque MN, Mebedi HMH, Rob MA, Salauddin A, et al. Dysbiosis of oral and gut microbiomes in SARS-CoV-2 infected patients in Bangladesh: elucidating the role of opportunistic gut microbes. *Front Med (Lausanne)* 2022; 9: 821777.
- Herrera D, Serrano J, Roldán S, Sanz M. Is the oral cavity relevant in SARS-CoV-2 pandemic? *Clin Oral Investig* 2020; 24(8): 2925–30.
- Zhao H, Chen S, Yang F, Wu H, Ba Y, Cui L, et al. Alternation of nasopharyngeal microbiota in healthy youth is associated with environmental factors: implication for respiratory diseases. *Int J Environ Health Res* 2022; 32(5): 952–62.
- Iebba V, Zanotta N, Campisciano G, Zerbato V, Di Bella S, Cason C, et al. Profiling of oral microbiota and cytokines in COVID-19 patients. *Front Microbiol* 2021; 12: 671813.
- Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. *JAMA* 2020; 323(11): 1061–9.
- Barros MIOS, Firmino GLO, Vieira T da S, Araujo A de A, Souza L de LB, Reis BS, et al. Oral manifestations associated with COVID-19 infection. *RSD [Internet]* 2021; [accessed on 19 Mar 2023] 10(16): e555101624107. Available from: <https://rsdjournal.org/index.php/rsd/article/view/24107>
- Haran JP, Bradley E, Zeamer AL, Cincotta L, Salive MC, Dutta P, et al. Inflammation-type dysbiosis of the oral microbiome associates with the duration of COVID-19 symptoms and long COVID. *JCI Insight* 2021; 6(20): e152346.
- Ward DV, Bhattarai S, Rojas-Correa M, Purkayastha A, Holler D, Qu MD, et al. The intestinal and oral microbiomes are robust predictors of COVID-19 severity the main predictor of COVID-19-related fatality [preprint]. 2021; medRxiv: 2021.01.05.20249061. Available from: <https://doi.org/doi:10.1101/2021.01.05.20249061>
- Singh K, Rao A. Probiotics: a potential immunomodulator in COVID-19 infection management. *Nutr Res* 2021; 87: 1–12.
- Ursell LK, Metcalf JL, Parfrey LW, Knight R. Defining the human microbiome. *Nutr Rev* 2012; 70 (Suppl 1): S38–44.
- Santacroce L, Charitos LA, Ballini A, Inchingolo F, Luperto P, De Nitto E, et al. The human respiratory system and its microbiome at a glimpse. *Biology (Basel)* 2020; 9(10): 318.
- Santacroce L, Sardaro N, Topi S, Pettini F, Bottalico L, Cantore S, et al. The pivotal role of oral microbiota in health and disease. *J Biol Regul Homeost Agents* 2020; 34(2): 733–7.
- Gomez P-AM. Use of probiotics in dentistry. *Dent Oral Craniofac Res* 2017; 4(1): 1–4.
- Lamont RJ, Koo H, Hajishengallis G. The oral microbiota: dynamic communities and host interactions. *Nat Rev Microbiol* 2018; 16(12): 745–59.
- Caselli E, Fabbri C, D'Acolti M, Soffritti I, Bassi C, Mazzavane S, et al. Defining the oral microbiome by whole-genome sequencing and resistome analysis: the complexity of the healthy picture. *BMC Microbiol* 2020; 20(1): 120.
- Radaic A, Kapila YL. The oralome and its dysbiosis: new insights into oral microbiome-host interactions. *Comput Struct Biotechnol J* 2021; 19: 1335–60.
- Wade WG. The oral microbiome in health and disease. *Pharmacol Res* 2013; 69(1): 137–43.
- Rowan-Nash AD, Korry BJ, Mylonakis E, Belenky P. Cross-domain and viral interactions in the microbiome. *Microbiol Mol Biol Rev* 2019; 83(1): e00044–18.
- Yamamoto S, Saito M, Tamura A, Pravisuda D, Mizutani T, Yotsuyanagi H. The human microbiome and COVID-19: a systematic review. *PLoS One* 2021; 16(6): e0253293.
- Biagi E, Candela M, Fairweather-Tait S, Franceschi C, Brigidi P. Aging of the human metaorganism: the microbial counterpart. *Age (Dordr)* 2012; 34(1): 247–67.
- Park SH, Kim KA, Ahn YT, Jeong JJ, Hub CS, Kim DH. Comparative analysis of gut microbiota in elderly people of urbanized towns and longevity villages. *BMC Microbiol* 2015; 15: 49.
- Arweiler NB, Netuschil L. The oral microbiota. *Adv Exp Med Biol* 2016; 902: 45–60.
- Brooks AW, Priya S, Blekhan R, Bordenstein SR. Gut microbiota diversity across ethnicities in the United States. *PLoS Biol* 2018; 16(12): e2006842.

40. Rinninella E, Raoul P, Cintoni M, Franceschi F, Miggiaro GAD, Gasbarrini A, et al. What is the healthy gut microbiota composition? A changing ecosystem across age, environment, diet, and diseases. *Microorganisms* 2019; 7(1): 14.
41. Liu J, Labousse L, Nivard MG, Bot M, Chen L, van Klinken JB, et al. Integration of epidemiologic, pharmacologic, genetic and gut microbiome data in a drug-metabolite atlas. *Nat Med* 2020; 26(1): 110–7.
42. Boyajian JL, Ghebretatios M, Schaly S, Islam P, Prakash S. Microbiome and human aging: probiotic and prebiotic potentials in longevity, skin health and cellular senescence. *Nutrients* 2021; 13(12): 4550.
43. Pérez-Brocá V, Moya A. The analysis of the oral DNA virome reveals which viruses are widespread and rare among healthy young adults in Valencia (Spain). *PLoS One* 2018; 13(2): e0191867.
44. Baker JL, Bor B, Agnello M, Shi W, He X. Ecology of the oral microbiome: beyond bacteria. *Trends Microbiol* 2017; 25(5): 362–74.
45. de la Cruz Peña MJ, Martínez-Hernández F, García-Heredia I, Lluésma Gomez M, Fornas O, Martínez-García M. Deciphering the human virome with single-virus genomics and metagenomics. *Viruses* 2018; 10(3): 113.
46. Peters BA, Wu J, Hayes RB, Ahn J. The oral fungal mycobiome: characteristics and relation to periodontitis in a pilot study. *BMC Microbiol* 2017; 17(1): 157.
47. Mosaddad SA, Tahmasebi E, Yazdani A, Rezvani MB, Seifalian A, Yazdani M, et al. Oral microbial biofilms: an update. *Eur J Clin Microbiol Infect Dis* 2019; 38(11): 2005–19.
48. Dubar M, Zaffino ML, Remen T, Thilly N, Cunat L, Machouart MC, et al. Protozoans in subgingival biofilm: clinical and bacterial associated factors and impact of scaling and root planing treatment. *J Oral Microbiol* 2019; 12(1): 1693222.
49. Belmok A, de Cena JA, Kyaw CM, Damé-Teixeira N. The oral archaeome: a scoping review. *J Dent Res* 2020; 99(6): 630–43.
50. Lu M, Xuan S, Wang Z. Oral microbiota: A new view of body health. *Food Sci Hum Wellness* 2019; 8(1): 8–15.
51. Stubbendieck RM, May DS, Chevette MG, Temkin MI, Wendt-Pienkowski E, Cagnazzo J, et al. Competition among nasal bacteria suggests a role for siderophore-mediated interactions in shaping the human nasal microbiota. *Appl Environ Microbiol* 2019; 85(10): e02406–18.
52. Gebrayel P, Nicco C, Al Khodor S, Bilinski J, Caselli E, Comelli EM, et al. Microbiota medicine: towards clinical revolution. *J Transl Med* 2022; 20(1): 111.
53. Edwards V, Smith DL, Meylan F, Tiffany L, Poncet S, Wu WW, et al. Analyzing the role of gut microbiota on the onset of autoimmune diseases using TNF^{ΔARE} murine model. *Microorganisms* 2021; 10(1): 73.
54. Petersen C, Round JL. Defining dysbiosis and its influence on host immunity and disease. *Cell Microbiol* 2014; 16(7): 1024–33.
55. Nikolić-Jakoba N, Vojnović S, Pavić A, Janković S, Leković V, Vasiljević B. Polymerase chain reaction in the identification of periodontopathogens: a reliable and satisfactory method? *Arch Biol Sci* 2012; 64(4): 1413–23.
56. Tandellin RT, Widita E, Agustina D, Saini R. The effect of oral probiotic consumption on the caries risk factors among high-risk caries population. *J Int Oral Health* 2018; 10(3): 132–7.
57. Zaura E, Tvetman S. Critical appraisal of oral pre- and probiotics for caries prevention and care. *Caries Res* 2019; 53(5): 514–26. <https://doi.org/10.1159/000499037>. PMID: 30947169
58. Ayala LDO, Zambrano JFB, Vire JMY, Gavilanes MPP, Coyago M de LR. Modulation of oral biofilm and immune response associated to mucosa with probiotic bacteria as a potential approach in the prevention of dental caries: a systematic review. *Dent Oral Biol Craniofac Res* 2020; 3(5): 1–7.
59. Talaat D. Effect of probiotic chewable tablets on oral health and white spot lesions in pre-school children: a randomized clinical trial. *Egypt Dent J* 2021; 67(3): 1797–807.
60. Miličević R, Brajović G, Nikolić-Jakoba N, Popović B, Pavlica D, Leković V, et al. Identification of periodontopathogen microorganisms by PCR technique. *Srp Arh Celok Lek* 2008; 136(9–10): 476–80. (Serbian)
61. Kumar PS. From focal sepsis to periodontal medicine: a century of exploring the role of the oral microbiome in systemic disease. *J Physiol* 2017; 595(2): 465–76.
62. Allaker RP, Stephen AS. Use of probiotics and oral health. *Curr Oral Health Rep* 2017; 4(4): 309–18.
63. Radović N, Nikolić-Jakoba N, Petrović N, Milosavljević A, Brković B, Roganović J. MicroRNA-146a and microRNA-155 as novel crevicular fluid biomarkers for periodontitis in nondiabetic and type 2 diabetic patients. *J Clin Periodontol* 2018; 45(6): 663–71.
64. Ratna Sudha M, Neelamraju J, Surendra Reddy M, Kumar M. Evaluation of the effect of probiotic *Bacillus coagulans* unique IS2 on mutans Streptococci and Lactobacilli levels in saliva and plaque: a double-blind, randomized, placebo-controlled study in children. *Int J Dent* 2020; 2020: 8891708.
65. Wang L, Ganly I. The oral mycrobioome and oral cancer. *Clin Lab Med* 2014; 34(4): 711–9.
66. Li Y, He J, He Z, Zhou Y, Yuan M, Xu X, et al. Phylogenetic and functional gene structure shifts of the oral microbiomes in periodontitis patients. *ISME J* 2014; 8(9): 1879–91.
67. Abusleme L, Morandini AC, Hasbişume-Takizawa T, Sabingur SE. Editorial: Oral microbiome and inflammation connection to systemic health. *Front Cell Infect Microbiol* 2021; 11: 780182.
68. Qin J, Li Y, Cai Z, Li S, Zhu J, Zhang F, et al. A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature* 2012; 490(7418): 55–60.
69. Marchi-Alves LM, Freitas D, de Andrade D, de Godoy S, Toneti AN, Mendes LAC. Characterization of oral microbiota in removable dental prosthesis users: influence of arterial hypertension. *Biomed Res Int* 2017; 2017: 3838640.
70. Bryan NS, Tribble G, Angelov N. Oral microbiome and nitric oxide: the missing link in the management of blood pressure. *Curr Hypertens Rep* 2017; 19(4): 33.
71. Cobb CM, Kelly PJ, Williams KB, Babbar S, Angolkar M, Derman RJ. The oral microbiome and adverse pregnancy outcomes. *Int J Womens Health* 2017; 9: 551–9.
72. Chen C, Hemme C, Beleno J, Shi ZJ, Ning D, Qin Y, et al. Oral microbiota of periodontal health and disease and their changes after nonsurgical periodontal therapy. *ISME J* 2018; 12(5): 1210–24.
73. Kaul D, Rathnasinghe R, Ferrer M, Tan GS, Barrera A, Pickett BE, et al. Microbiome disturbance and resilience dynamics of the upper respiratory tract during influenza A virus infection. *Nat Commun* 2020; 11(1): 2537.
74. Lee HS, Lobbstaël E, Vermeire S, Sabino J, Cleynen I. Inflammatory bowel disease and Parkinson's disease: common pathophysiological links. *Gut* 2021; 70(2): 408–17.
75. Willmott T, McBain AJ, Humphreys GJ, Myers J, Cottrell E. Does the oral microbiome play a role in hypertensive pregnancies? *Front Cell Infect Microbiol* 2020; 10: 389.
76. Sobail MU, Hedin L, Al-Asmakb M. Dysbiosis of the salivary microbiome is associated with hypertension and correlated with metabolic syndrome biomarkers. *Diabetes Metab Syndr Obes* 2021; 14: 4641–53.
77. Milasin J, Nikolić-Jakoba N, Stefanović D, Sopta J, Pucar A, Leković V, et al. Periodontal inflammation as risk factor for pancreatic diseases. In: Nagal A, editor. *Inflammatory diseases - a modern perspective*. London: IntechOpen; 2011.
78. Dordević V, Jovanović M, Stefanović V, Nikolić-Jakoba N, Đokić G, Stašević Karličić I, et al. Assessment of periodontal health among

- the inpatients with schizophrenia. *Vojnosanit Pregl* 2019; 76(11): 1139–46.
79. Ren Z, Wang H, Cui G, Lu H, Wang L, Luo H, et al. Alterations in the human oral and gut microbiomes and lipidomics in COVID-19. *Gut* 2021; 70(7): 1253–65.
 80. Zhang X, Zhang D, Jia H, Feng Q, Wang D, Liang D, et al. The oral and gut microbiomes are perturbed in rheumatoid arthritis and partly normalized after treatment. *Nat Med* 2015; 21(8): 895–905.
 81. Kurt BS, Ilhan B, Sevki BI, Kurt AF, Orhan K. Periodontal management during COVID-19 pandemic: mini review. *Balk J Dent Med* 2021; 25(3): 135–8.
 82. Rakić M, Nikolić-Jakoba N, Struillout X, Petković-Čurčin A, Stamatović N, Matić S, et al. Receptor activator of nuclear factor kappa B (RANK) as a determinant of peri-implantitis. *Vojnosanit Pregl* 2013; 70(4): 346–51.
 83. Kilian M, Chapple IL, Hannig M, Marsh PD, Meuric V, Pedersen AM, et al. The oral mycobiome - an update for oral healthcare professionals. *Br Dent J* 2016; 221(10): 657–66.
 84. Belkaid Y, Hand TW. Role of the microbiota in immunity and inflammation. *Cell* 2014; 157(1): 121–41.
 85. Trompette A, Gollwitzer ES, Pattaroni C, Lopez-Mejia IC, Riva E, Pernot J, et al. Dietary fiber confers protection against flu by shaping Ly6c⁺ patrolling monocyte hematopoiesis and CD8⁺ T cell metabolism. *Immunity* 2018; 48(5): 992–1005. e8.
 86. Willis JR, Gabaldón T. The human oral mycobiome in health and disease: from sequences to ecosystems. *Microorganisms* 2020; 8(2): 308.
 87. Lynch SV. Viruses and microbiome alterations. *Ann Am Thorac Soc* 2014; 11(Suppl 1): S57–60.
 88. Li N, Ma WT, Pang M, Fan QL, Hua JL. The commensal microbiota and viral infection: a comprehensive review. *Front Immunol* 2019; 10: 1551.
 89. Scannapieco FA. Role of oral bacteria in respiratory infection. *J Periodontol* 1999; 70(7): 793–802.
 90. Khan R, Petersen FC, Shekbar S. Commensal Bacteria: An emerging player in defense against respiratory pathogens. *Front Immunol* 2019; 10: 1203.
 91. Pfeiffer JK, Sonnenburg JL. The intestinal microbiota and viral susceptibility. *Front Microbiol* 2011; 2: 92.
 92. Hanada S, Pirzadeh M, Carver KY, Deng JC. Respiratory viral infection-induced microbiome alterations and secondary bacterial pneumonia. *Front Immunol* 2018; 9: 2640.
 93. Ai JW, Zhang HC, Xu T, Wu J, Zhu M, Yu YQ, et al. Optimizing diagnostic strategy for novel coronavirus pneumonia, a multi-center study in Eastern China [preprint]. 2020; medRxiv: 2020.02.13.20022673. Available from: <https://www.medrxiv.org/content/10.1101/2020.02.13.20022673v1>
 94. Xu K, Cai H, Shen Y, Ni Q, Chen Y, Hu S, et al. Management of COVID-19: the Zhejiang experience. *Zhejiang Da Xue Xue Bao Yi Xue Ban* 2020; 49(2): 147–57. (Chinese)
 95. Cox MJ, Loman N, Bogaert D, O'Grady J. Co-infections: potentially lethal and unexplored in COVID-19. *Lancet Microbe* 2020; 1(1): e11.
 96. Roncati L, Lusenti B, Nasillo V, Manenti A. Fatal SARS-CoV-2 coinfection in course of EBV-associated lymphoproliferative disease. *Ann Hematol* 2020; 99(8):1945–6.
 97. Chakraborty S. Metagenome of SARS-Cov-2 patients in Shenzhen with travel to Wuhan shows a wide range of species - *Lautropia*, *Cutibacterium*, *Haemophilus* being most abundant - and *Campylobacter* explaining diarrhea [preprint]. *OSF Preprints* 2020; doi: 10.31219/osf.io/jegwq.
 98. Wu F, Zhao S, Yu B, Chen YM, Wang W, Song ZG, et al. A new coronavirus associated with human respiratory disease in China. *Nature* 2020; 579(7798): 265–9. Erratum in: *Nature* 2020; 580(7803): E7.
 99. Chen T, Song J, Liu H, Zheng H, Chen C. Positive Epstein-Barr virus detection in coronavirus disease 2019 (COVID-19) patients. *Sci Rep* 2021; 11(1): 10902.
 100. Yeob YK, Zuo T, Lui GC, Zhang F, Liu Q, Li AY, et al. Gut microbiota composition reflects disease severity and dysfunctional immune responses in patients with COVID-19. *Gut* 2021; 70(4): 698–706.
 101. Marouf N, Cai W, Said KN, Daas H, Diab H, Chinta VR, et al. Association between periodontitis and severity of COVID-19 infection: a case-control study. *J Clin Periodontol* 2021; 48(4): 483–91.
 102. Song WJ, Hui CKM, Hull JH, Birring SS, McGarvey L, Mazzone SB, et al. Confronting COVID-19-associated cough and the post-COVID syndrome: role of viral neurotropism, neuroinflammation, and neuroimmune responses. *Lancet Respir Med* 2021; 9(5): 533–44.
 103. Oronsky B, Larson C, Hammond TC, Oronsky A, Kesari S, Lybeck M, et al. A review of persistent post-COVID syndrome (PPCS). *Clin Rev Allergy Immunol* 2023; 64(1): 66–74; Epub 2021; 20: 1–9.
 104. France K, Glick M. Long COVID and oral health care considerations. *J Am Dent Assoc* 2022; 153(2): 167–74.
 105. Larsen JM. The immune response to *Prevotella* bacteria in chronic inflammatory disease. *Immunology* 2017; 151(4): 363–74.
 106. Xu L, Chen X, Wang Y, Jiang W, Wang S, Ling Z, et al. Dynamic alterations in salivary microbiota related to dental caries and age in preschool children with deciduous dentition: a 2-year follow-up study. *Front Physiol* 2018; 9: 342.
 107. Khan AA, Khan Z. COVID-2019-associated overexpressed *Prevotella* proteins mediated host-pathogen interactions and their role in coronavirus outbreak. *Bioinformatics* 2020; 36(13): 4065–9.
 108. Wu Y, Cheng X, Jiang G, Tang H, Ming S, Tang L, et al. Author correction: Altered oral and gut microbiota and its association with SARS-CoV-2 viral load in patients with COVID-19 during hospitalization. *NPJ Biofilms Microbiomes* 2021; 7(1): 90. Erratum for: *NPJ Biofilms Microbiomes* 2021; 7(1): 61.
 109. Jasinski-Bergner S, Mandelboim O, Seliger B. Molecular mechanisms of human herpes viruses inferring with host immune surveillance. *J Immunother Cancer* 2020; 8(2): e000841.
 110. Azarpazhoob A, Leake JL. Systematic review of the association between respiratory diseases and oral health. *J Periodontol* 2006; 77(9): 1465–82.
 111. Hols P, Ledesma-García L, Gabant P, Mignolet J. Mobilization of microbiota commensals and their bacteriocins for therapeutics. *Trends Microbiol* 2019; 27(8): 690–702.
 112. Barbour A, Wescombe P, Smith L. Evolution of Lantibiotic Salivaricins: New weapons to fight infectious diseases. *Trends Microbiol* 2020; 28(7): 578–93.
 113. Hadžić Z, Pašić E, Gojko-Vukelić M, Hadžić S. Effects of *Lactobacillus reuteri* lozenges (Prodentis) as adjunctive therapeutic agent in non-surgical therapy of periodontitis. *Balk J Dent Med* 2021; 25(1): 41–5.
 114. Bottari B, Castellone V, Neviani E. Probiotics and Covid-19. *Int J Food Sci Nutr* 2021; 72(3): 293–9.
 115. Mikulić A, Bakarić D, Ivanić Jokić N, Hrvatinić S, Culav T. The use of probiotics in dental medicine. *Madridge J Dent Oral Surg* 2017; 2(1): 44–6.
 116. Bermudez-Brito M, Plaza-Díaz J, Muñoz-Quezada S, Gómez-Llrente C, Gil A. Probiotic mechanisms of action. *Ann Nutr Metab* 2012; 61(2): 160–74.
 117. Eguchi K, Fujitani N, Nakagawa H, Miyazaki T. Prevention of respiratory syncytial virus infection with probiotic lactic acid

- bacterium *Lactobacillus gasseri* SBT2055. *Sci Rep* 2019; 9(1): 4812.
118. *Morawej H, Memariani H, Memariani M.* Therapeutic and preventive potential of probiotics against COVID-19. *Res Bull Med Sci* 2020; 25(1): e18.
 119. *Azad MAK, Sarker M, Wan D.* Immunomodulatory effects of probiotics on cytokine profiles. *Biomed Res Int* 2018; 2018: 8063647.
 120. *Hardy H, Harris J, Lyon E, Beal J, Foey AD.* Probiotics, prebiotics and immunomodulation of gut mucosal defences: homeostasis and immunopathology. *Nutrients* 2013; 5(6): 1869–912.
 121. *Al Kassaa I.* The antiviral activity of probiotic metabolites. In: *Al Kassaa I.* New insights on antiviral probiotics. Springer, Cham: 2017; 83–97.
 122. *Xia Y, Cao J, Wang M, Lu M, Chen G, Gao F, et al.* Effects of *Lactococcus lactis* subsp. *lactis* JCM5805 on colonization dynamics of gut microbiota and regulation of immunity in early ontogenetic stages of tilapia. *Fish Shellfish Immunol* 2019; 86: 53–63.
 123. *Hung YP, Lee CC, Lee JC, Tsai PJ, Ko WC.* Gut dysbiosis during COVID-19 and potential effect of probiotics. *Microorganisms* 2021; 9(8): 1605.
 124. *Al Kassaa I, Hober D, Hamze M, Chibib NE, Drider D.* Antiviral potential of lactic acid bacteria and their bacteriocins. *Probiotics Antimicrob Proteins* 2014; 6(3–4): 177–85.
 125. *Mabooti M, Abdolalipour E, Salehzadeh A, Mobebe SR, Gorji A, Ghaemi A.* Immunomodulatory and prophylactic effects of *Bifidobacterium bifidum* probiotic strain on influenza infection in mice. *World J Microbiol Biotechnol* 2019; 35(6): 91.
 126. *Baindara P, Chakraborty R, Holliday ZM, Mandal SM, Sebrum AG.* Oral probiotics in coronavirus disease 2019: connecting the gut-lung axis to viral pathogenesis, inflammation, secondary infection and clinical trials. *New Microbes New Infect* 2021; 40: 100837.
 127. *Malaczewska J, Kaczorek-Lukowska E, Wójcik R, Sivnicki AK.* Antiviral effects of nisin, lysozyme, lactoferrin and their mixtures against bovine viral diarrhoea virus. *BMC Vet Res* 2019; 15(1): 318.
 128. *Di Pierro F.* A possible probiotic (*S. salivarius* K12) approach to improve oral and lung microbiotas and raise defenses against SARS-CoV-2. *Minerva Med* 2020; 111(3): 281–3.
 129. *Zhang H, Yeh C, Jin Z, Ding L, Liu BY, Zhang L, et al.* Prospective study of probiotic supplementation results in immune stimulation and improvement of upper respiratory infection rate. *Synth Syst Biotechnol* 2018; 3(2): 113–20.
 130. *Seminario-Amez M, López-López J, Estrugo-Devesa A, Ayuso-Montero R, Jané-Salas E.* Probiotics and oral health: a systematic review. *Med Oral Patol Oral Cir Bucal* 2017; 22(3): e282–8.
 131. *Conte L, Toraldo DM.* Targeting the gut-lung microbiota axis by means of a high-fibre diet and probiotics may have anti-inflammatory effects in COVID-19 infection. *Ther Adv Respir Dis* 2020; 14: 1753466620937170.
 132. *Olaimat AN, Aolymat I, Al-Holy M, Ayyash M, Abu Ghoush M, Al-Nabulsi AA, et al.* The potential application of probiotics and prebiotics for the prevention and treatment of COVID-19. *NPJ Sci Food* 2020; 4: 17.
 133. *Stavropoulou E, Beziirtzoglou E.* Probiotics in medicine: a long debate. *Front Immunol* 2020; 11: 2192.
 134. *Xavier-Santos D, Padilha M, Fabiano GA, Vinderola G, Gomes Cruz A, Sivieri K, et al.* Evidences and perspectives of the use of probiotics, prebiotics, synbiotics, and postbiotics as adjuvants for prevention and treatment of COVID-19: a bibliometric analysis and systematic review. *Trends Food Sci Technol* 2022; 120: 174–92. Erratum in: *Trends Food Sci Technol* 2022; 121: 156–160.
 135. *d'Ettore G, Ceccarelli G, Marazzato M, Campagna G, Pinacchio C, Alessandri F, et al.* Challenges in the management of SARS-CoV2 infection: the role of oral bacteriotherapy as complementary therapeutic strategy to avoid the progression of COVID-19. *Front Med (Lausanne)* 2020; 7: 389.
 136. *Tang H, Bobannon L, Lew M, Jensen D, Jung SH, Zhao A, et al.* Randomised, double-blind, placebo-controlled trial of probiotics to eliminate COVID-19 transmission in exposed household contacts (PROTECT-EHC): a clinical trial protocol. *BMJ Open* 2021; 11(5): e047069.
 137. *Endam LM, Tremblay C, Filali A, Desrosiers MY.* Intranasal application of *Lactococcus lactis* W136 bacteria early in SARS-CoV-2 infection may have a beneficial immunomodulatory effect: A proof-of-concept study [preprint]. 2021; medRxiv: 2021.01.05.20249061. Available from: <https://www.medrxiv.org/content/10.1101/2021.04.18.21255699v1.full>
 138. *Gutiérrez-Castrellon P, Gandara-Martí T, Abreu AT, Nieto-Rujfno CD, López-Orduna E, Jimenez-Escobar I, et al.* Efficacy and safety of novel probiotic formulation in adult Covid19 outpatients: a randomized, placebo-controlled clinical trial [preprint]. 2021; medRxiv: 2021.05.20.21256954. Available from: <https://www.medrxiv.org/content/10.1101/2021.05.20.21256954v1.full.pdf+html>
 139. *Mozgata M, Castro I, Gomez-Torres N, Arroyo R, Lailla Y, Somada M, et al.* Administration of *Ligilactobacillus salivarius* MP101 in an elderly nursing home during the COVID-19 pandemic: immunological and nutritional impact. *Foods* 2021; 10(9): 2149.
 140. *Wang Q, Lin X, Xiang X, Liu W, Fang Y, Chen H, et al.* Oropharyngeal probiotic ENT-K12 prevents respiratory tract infections among frontline medical staff fighting against COVID-19: a pilot study. *Front Bioeng Biotechnol* 2021; 9: 646184.
 141. *Li M, He Z, Yang J, Guo Q, Weng H, Luo J, et al.* Clinical characteristics, outcomes, and risk factors of disease severity in patients with COVID-19 and with a history of cerebrovascular disease in Wuhan, China: A Retrospective Study. *Front Neurol* 2022; 12: 706478.
 142. *Bozkurt HS, Bilen Ö.* Oral booster probiotic bifidobacteria in SARS-COV-2 patients. *Int J Immunopathol Pharmacol* 2021; 35: 20587384211059677.
 143. *Wischmeyer PE, Tang H, Ren Y, Bobannon L, Ramirez ZE, Andermann TM, et al.* Daily lactobacillus probiotic versus placebo in COVID-19-exposed household contacts (PROTECT-EHC): a randomized clinical trial [preprint]. 2022; medRxiv: 2022.01.04.21268275. Available from: <https://www.medrxiv.org/content/10.1101/2022.01.04.21268275v1>
 144. *Block J.* High risk COVID-19: potential intervention at multiple points in the COVID-19 disease process via prophylactic treatment with azithromycin or bee derived products. *J Biomed Res Rev* 2020; 3(1): 26–31.
 145. *Horowitz RI, Freeman PR, Bruzzese J.* Efficacy of glutathione therapy in relieving dyspnea associated with COVID-19 pneumonia: a report of 2 cases. *Respir Med Case Rep* 2020; 30: 101063.
 146. *Pourbossein M, Moravejolahkami AR.* Probiotics in viral infections, with a focus on COVID-19: a systematic review [preprint]. Authorea 2020; doi: 10.22541/au.158999387.76467979.
 147. *Ceccarelli G, Borrazzo C, Pinacchio C, Santinelli L, Innocenti GP, Cavallari EN, et al.* Oral bacteriotherapy in patients with COVID-19: a retrospective cohort study. *Front Nutr* 2021; 7: 613928.
 148. *Patra S, Saxena S, Sahu N, Pradhan B, Roychowdhury A.* Systematic network and meta-analysis on the antiviral mechanisms of probiotics: a preventive and treatment strategy to mitigate SARS-CoV-2 infection. *Probiotics Antimicrob Proteins* 2021;13(4): 1138–56.

149. *Ayyash M, Olaimat A, Al-Nabulsi A, Liu SQ.* Bioactive properties of novel probiotic *Lactococcus lactis* fermented camel sausages: cytotoxicity, angiotensin converting enzyme inhibition, antioxidant capacity, and antidiabetic activity. *Food Sci Anim Resour* 2020; 40(2): 155–71.
150. *Nayebi A, Navashenaq JG, Soleimani D, Nachvak SM.* Probiotic supplementation: a prospective approach in the treatment of COVID-19. *Nutr Health* 2022; 28(2): 163–75.
151. *Fernández-Fernández FJ.* COVID-19, hypertension and angiotensin receptor-blocking drugs. *J Hypertens* 2020; 38(6): 1191.
152. *Imai Y, Kuba K, Rao S, Huan Y, Guo F, Guan B,* et al. Angiotensin-converting enzyme 2 protects from severe acute lung failure. *Nature* 2005; 436(7047): 112–6.
153. *Spagnolello O, Pinacchio C, Santinelli L, Vassalini P, Innocenti GP, De Girolamo G,* et al. Targeting microbiome: an alternative strategy for fighting SARS-CoV-2 infection. *Chemotherapy* 2021; 66(1–2): 24–32.
154. *Infusino F, Marazzato M, Mancone M, Fedele F, Mastroianni CM, Severino P,* et al. Diet supplementation, probiotics, and nutraceuticals in SARS-CoV-2 infection: a scoping review. *Nutrients* 2020; 12(6): 1718.
155. *Santos TGFTD, Brito DHS, Santos NMVD, Paiva MC, Lyra MCA, Heimer MV,* et al. Viral symptoms in children and SARS-COV-2: information for pediatric dentists for the control of transmission. *Braz Oral Res* 2022; 36: e029.

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